Monatshefte für Chemie Chemical Monthly © Springer-Verlag 1998 Printed in Austria

Concerning the Dissociation of Hypericin: An Atmospheric Pressure Ionization Mass Spectrometric Study

Werner Ahrer, Heinz Falk * , and Huyen T. N. Tran

Institut für Chemie, Johannes Kepler University A-4040 Linz, Austria

Summary. The dissociation of hypericin (1) was investigated using atmospheric pressure ionization mass spectrometry of 80% ethanolic solutions under variation of their pH values. Thus, it could be unequivocally established that the species predominant within the pH region of 3–10 and 11–13 are the mono- and dideprotonated compounds $1^{(-)}$ and $1^{(2-)}$. These results nicely corroborate the hitherto accumulated mostly spectrophotometric evidence for the first deprotonation step occurring at a $pK₂$ value in the order of 2 and the second one at about 12. Moreover, this example demonstrates the potential of atmospheric pressure ionization mass spectrometry as a valuable tool to investigate dissociation equilibria.

Keywords. Deprotonation; Hypericin; Atmospheric pressure ionization mass spectrometry; pK_a values.

Zur Dissoziation des Hypericins: Eine massenspektrometrische Untersuchung mit Atmosphärendruck-Ionisierung

Zusammenfassung. Die Dissoziation von Hypericin (1) wurde mit Hilfe der Elektrospray-Massenspektrometrie von Lösungen in 80% Ethanol unter Variation des pH Wertes untersucht. Damit konnte eindeutig abgeleitet werden, daû die vorherrschenden Spezies innerhalb der Bereiche von pH 3 -10 und 11 -13 mono- und dideprotoniert sind. Dieses Ergebnis bestätigt die bislang erzielte vornehmlich spektrophotometrische Evidenz für einen ersten Deprotonierungsschritt im pK_a -Bereich von 2 und eines zweiten bei etwa 12. Darüber hinaus demonstriert dieses Beispiel das Potential der Atmosphärendruckionisations-Massenspektrometrie für die Untersuchung von Dissoziationsgleichgewichten.

Introduction

The natural pigment hypericin (1), contained mostly in Hypericum species [1], displays a variety of physiological effects [1, 2]. Several of them seem to depend critically upon its unique acid-base properties [3]. Since the protonationdeprotonation behavior of 1 is thought to be of fundamental importance for its

Corresponding author

physiological effects, there is a need to characterize the various species present in aqueous systems.

Investigations have been undertaken to derive pK_a values of 1 by means of ¹H NMR and UV/Vis spectroscopy and electrophoresis, especially with respect to its physiologically meaningful deprotonation steps [4, 5]. However, such studies have been severely limited by the notorious solubility problems encountered with 1. Remedies have been sought by using certain derivatives, like ω, ω' -oligoethyleneglycol conjugates, for such measurements [5]. However, the results of these studies as shown in Scheme 1 for water as the solvent (bay denotes the 3/4- and peri the 1/6/8/13-positions), mainly with regard to the species involved, have not yet been generally accepted. Therefore, any additional methodologically independent experimental evidence brought about, could be of value.

$$
1 = OH^{+} \xrightarrow{pK_{a} = -6} 1 \xrightarrow{pK_{a} = 1.8} 1_{\text{bay}} - O^{-} \xrightarrow{pK_{a} = 9.2} 1_{\text{bay}} - O^{-}, \text{peri} - O^{-}
$$

Scheme 1

Ionization in the course of atmospheric pressure ionization mass spectrometry forms intact, low internal energy, even-electron molecular ions by protonation or deprotonation of neutral molecules having acidic and/or basic functional groups, depending also on the acid or base properties of the medium [6]. Thus, it could be envisaged that upon spraying 1 from solvents of different pH values, molecular ions would result that are characteristic for their state of dissociation in the bulk medium. This report will describe such experiments.

Results and Discussion

Recording the atmospheric pressure ionization mass spectrum of 1 dissolved in an 80% ethanol water mixture ($pH \approx 5$) yielded a spectrum with essentially the molecular ion peak of the monodeprotonated system $1^{(-)}$ at $m/z = 503$ $(M_{1calc} = 504)$. Upon lowering the pH value of the solution this ion persisted down to about pH 3. From pH 3.5 downwards, 1 started to precipitate, and no ions involving 1 but exclusively ions characteristic of the acid used for acidification (HCl, H₃PO₄) were observed, e.g. $3H_3O^+ \cdot 3Cl^-$, $m/z = 162$; $3H_3O^+ \cdot 3Cl^- \cdot HCl$, $m/z = 198; H_3PO_4 \cdot H_2PO_4$, $m/z = 195; 2H_3PO_4 \cdot H_2PO_4$, $m/z = 293;$ $3H_3PO_4 \cdot H_2PO_4^-$, $m/z = 391$; $4H_3PO_4 \cdot H_2PO_4^-$, $m/z = 489$; $5H_3PO_4 \cdot H_2PO_4^-$, $m/z = 55$ 587. The relative abundance of the $1^{(-)}$ ion varied with the solution's pH value as illustrated in Fig. 1. This behavior is consistent with a pK_a value for the first deprotonation step of 1.7 of 1 dissolved in 80% ethanol [7] and of 1.5 in 80%

Fig. 1. Atmospheric pressure ionization mass spectrometric relative abundances (normalized to their maximum peak intensity) of the hypericinate ions $1^{(-)}$ (m/z = 503; —) and $1^{(2-)}$ (m/z = 251; \cdots) in dependence of the bulk pH values of a solution of 1 in 80% ethanol

DMSO/water [8], provided that the extremely low solubility of 1 in these acid systems is properly taken into account. Accordingly, rising precipitation in this p H range would lead (from an elaboration of Fig. 1) to an apparent pK_a value of about 4. This phenomenon of substantially higher apparent pK_a values is commonly observed for dissociation equilibria coupled to solubility equilibria, like in the case of the bilirubin pK_a values [9].

Of course, one has to be also aware of a recent study which showed that there could be a difference between the acidity of the bulk solution and the acidity within the atmospheric pressure ionization droplets [10]. However, this effect has been found to be large for neutral solutions, with an $10^3 - 10^4$ fold increase in acidity, but it has been found to be more or less negligible for already acidic solutions [10]. Thus, this effect could also contribute to the higher apparent pK_a value of 1, but the main reason for the difference between apparent and spectrophotometric pK_a values has to be blamed to the insolubility of 1. Nevertheless, the present atmospheric pressure ionization mass spectrometric experiment proved nicely, and beyond any doubt, that the species present in the pH range above 3 was the monodeprotonated hypericin $\mathbf{1}^{(-)}$. With respect to the position of deprotonation, a variety of experiments have shown it to be located at the bay region i.e. at position 3 [4, 5].

Upon increasing the pH value of the solution of 1 the relative abundance of the sole ion at $m/z = 503$ kept its intensity at a plateau value and then started to decrease at a pH value of about 10 to vanish around pH 14. Within this latter range a new ion was observed which was characterized by $m/z = 251$ (cf. Fig. 1). Its abundance peaked at about pH 13, and it had to be attributed $(m/z = 502/2 = 251)$ to the doubly charged dideprotonated species $1^{(2-)}$. The corresponding apparent pK_a value of about 12 as deduced from Fig. 1 was in accordance with the spectrophotometric titration values of 12.5 and 13 observed for 80% ethanol and 80% dimethylsulfoxide solutions [7, 8]. The species pre-dominant at pH values in the region of 13 was thus found from the mass spectroscopic data to be the dideprotonated hypericin ion $1^{(2-)}$, corroborating recent assignments [5, 8]. With respect to its structural details it has been deduced that the second deprotonation

step takes place at a peri position [5, 8]. It should also be stressed that from the difference between apparent and spectrophotometric pK_a values there seemed to be no significant difference between the bulk and spray droplet acidities in the basic region.

It should be mentioned that in the range of decreasing abundance of the $1^{(2-)}$ ion, *i.e.* at pH values of about 13–14, the trideprotonated hypericin $(1^{(3-)})$ at $m/z = 167$ could not be observed with certainty, since in contrast to the acidic and slightly basic conditions, where the molecular peak dominated the spectra, basic conditions led to a variety of mass fragments. However, the sharp decrease in the $1^{(2-)}$ ion abundance between pH 13 and 14 suggests that there is a further deprotonation step (cf. Fig. 1). This trideprotonation step has been hitherto observed only in the case of certain hypericin analogous compounds [8] to occur in the range characterized by pK_a values above 13.

Conclusions

Atmospheric pressure ionization mass spectrometry of solutions of hypericin (1) involving a series of pH values unequivocally demonstrated that the species present within the pH ranges of 3-10 and 11-13 are the mono- and dideprotonated ions $1^{(-)}$ and $1^{(2-)}$. These results nicely corroborate the hitherto accumulated evidence for the first deprotonation occurring at a pK_a value in the order of 2 and the second one at about 12 depending on the solvents used. With respect to the methodological aspect, the example of hypericin served to illustrate that atmospheric pressure ionization mass spectrometry under variation of pH constitutes a valuable tool to clarify the nature of species involved in dissociation equilibria.

Experimental

Hypericin (1) was prepared by means of the recently described semi-synthetic procedure from emodin which was extracted from the commercially available bark, Cortex frangulae [11].

Samples were prepared by saturating 1 in 80% ethanol water mixtures and adjusting the pH values by means of HCl or H_3PO_4 on the one hand, and by means of NaOH or NH₃ on the other hand. The pH values of the resulting samples were measured using a Metrohm-654 pH-meter equipped with a glass electrode.

Atmospheric pressure ionization mass spectrometry was performed in the negative ion mode using a quadrupole instrument (HP 5989B, Hewlett Packard, Palo Alto, CA) equipped with an Atmospheric Pressure Ionization interface (HP 59987A). Sample introduction was performed by continuous flow injection using a syringe pump (model 22, Harvard Apparatus, South Natick, MA) at a flow rate of 15 μ l/min. The spraying gas (N₂ 5.0) was delivered at a pressure of 60 psi, the drying gas (N₂ 5.0) was delivered at a flow of approximately 7 l/min and at a temperature of 150 °C. The voltages of the interface were set to $V_{\text{cyl}} = 3350 \text{ V}$, $V_{\text{end}} = 2580 \text{ V}$, and $V_{\text{cap}} = 3350 \text{ V}$. The electron multiplier voltage was adjusted to 1795 V.

Acknowledgments

We are grateful to Prof. Dr. W. Buchberger for generously providing access to the mass spectrometer. Sponsoring by the Fonds zur Förderung der Wissenschaftlichen Forschung (project No. P10680-CHE) is gratefully acknowledged.

References

- [1] Roth L (1990) Hypericum±Hypericin: Botanik. Inhaltsstoffe, Wirkung. ecomed, Landsberg
- [2] Song PS, Häder DP, Poff K (1980) Photochem Photobiol 32: 781; Giese AC (1981) Photochem Photobiol Rev 6: 139; Meruelo D, Lavie G, Lavie D (1988) Proc Natl Acad Sci USA 85: 5230; Lavie G, Valentine F, Levin B, Mazur Y, Gallo G, Lavie D, Weiner D, Meruelo D (1989) Proc Natl Acad USA 86: 5963; Tang J, Calacino JM, Larsen SH, Spitzer W (1990) Antiviral Research 13: 313; Kim I, Rhee J, Huh JW, Florell S, Faure B, Lee KW, Kahsai T, Song PS, Tamai N, Yamazaki T, Yamazaki I (1990) Biochim Biophys Acta 1040: 43; Lopez-Bazzocchi I, Hudson JB, Towers HN (1991) Photochem Photobiol 54: 95; Carpenter S, Kraus GA (1991) Photochem Photobiol 53: 169; Tao N, Orlando M, Hyon J, Grass M, Song PS (1993) J Am Chem Soc 115: 2526; Wynn JL, Kim JH, Tao N, Dai R, Song PS, Cotton TM (1995) J Phys Chem 99: 2208; Agostinis P, Donella-Deana A, Cuveele J, Vandenbogaerde A, Sarno S, Merlevede W, deWitte P (1996) Biochem Biophys Res Commun 220: 613; Kubin A, Alth G, Jindra RH, Jessner G, Ebermann R (1996) J Photochem Photobiol B 36: 103; Koren H, Schenk GM, Jindra RH, Alth G, Ebermann R, Kubin A, Koderhold G, Kreitner M (1996) J Photochem Photobiol B 36: 113
- [3] Fehr MJ, McCloskey MA, Petrich JW (1995) J Am Chem Soc 117: 1833; Fehr MJ, Carpenter SL, Wannemuehler Y, Petrich JW (1995) Biochemistry 34: 15845; Sureau F, Miskovsky P, Chinsky L, Turpin PY (1996) J Chem Soc 118: 9484
- [4] Etzlstorfer C, Falk H, Mayr E, Schwarzinger S (1996) Monatsh Chem 127: 1229
- [5] Altmann R, Falk H (1997) Monatsh Chem 128: 571
- [6] Burlingame AL, Boyd RK, Gaskell SJ (1996) Anal Chem 68: 599R; Hirabayashi A, Sakairi M, Takada Y, Koizumi H (1997) Trends Anal Chem 16: 45
- [7] Freeman D, Frolow F, Kapinus E, Lavie D, Lavie G, Meruelo D, Mazur Y (1994) J Chem Soc Chem Commun 891
- [8] Falk H, Mayr E (1995) Monatsh Chem 126: 699
- [9] Lightner DA, Holmes DL, McDonagh AF (1996) J Biol Chem 271: 2397
- [10] Gatlin CL, Turecek F (1994) Anal Chem 66: 712
- [11] Falk H, Meyer J, Oberreiter M (1993) Monatsh Chem 124: 339

Received December 31, 1997. Accepted (revised) January 30, 1998